

Low-density lipoprotein oxidation

Luigi Iuliano, Fausta Micheletta, Francesco Violi

Institute of Clinical Medicine I, University of Rome "La Sapienza", Rome, Italy

Key words:
Atherosclerosis;
Free radicals;
Low-density lipoproteins;
Oxidative stress.

Free radical mediated oxidation of low-density lipoproteins (LDL), which has been extensively studied in the last two decades, plays a central role in the development of the atherosclerotic plaque. Oxidation involves the lipid moiety of LDL in a chain reaction mechanism. In the initial phase, free radicals preferentially attack highly oxidizable polyunsaturated fatty acids. Subsequent recruitment of other molecules includes cholesterol and phospholipids. The process of oxidation is counteracted by antioxidants present in LDL. By-products formed during oxidation of LDL lipids, which may have biological activity, react with amino acid residues of the LDL protein backbone with the consequent modification of chemical and immunological properties responsible for cellular receptor shift. Oxidation-altered apolipoprotein B of oxidized LDL is, in fact, recognized by the macrophage scavenger receptor responsible for foam cell formation. The mechanism of LDL oxidation and the impact on atherogenesis are discussed.

(*Ital Heart J* 2001; 2 (12): 867-872)

© 2001 CEPI Srl

Address:

Prof. Francesco Violi
*Istituto di Clinica Medica
Università degli Studi
"La Sapienza"
Policlinico Umberto I
Viale del Policlinico, 155
00161 Roma
E-mail: francesco.violi@uniroma1.it*

Introduction

Atherosclerosis is a common disease in western population and plays a central role in the pathophysiology of cardiovascular disease. In the last two decades a series of excellent studies unraveled biochemical mechanisms which provided the background for a theory of atherogenesis. The backbone of this theory is based on lipid laden cells of atherosclerotic lesion (foam cells), on plasma lipoproteins, on a free radical-mediated modification of plasma lipoproteins, and on the scavenger receptor¹. Foam cells are the main cell type of atherosclerotic lesions and have been identified as differentiated from monocytes migrated from blood and from smooth muscle cells of the arterial wall²⁻⁴. Among plasma lipoproteins, the low-density lipoprotein (LDL) is the main protein responsible for transport and accumulation of lipids in the arterial wall. Paradoxically, accumulation of LDL in developing foam cells does not occur by the way of classic LDL receptor⁵. Incubation of macrophage with even very high concentrations of LDL does not increase cholesterol content, and patients with familial hypercholesterolemia who totally lack LDL receptors develop atherosclerotic plaques plenty of foam cells^{5,6}. LDL needs to be chemically altered/modified to enter the cells of atherosclerotic plaque via an unregulated receptor, the scavenger receptor⁵.

Low-density lipoprotein structure and metabolism

The metabolic precursor of LDL is the very low-density lipoprotein (VLDL) constructed in the liver and secreted into the circulation where becomes a mature LDL particle after lipid exchange and remodeling. Lipoprotein lipase, which is present on the surface of the endothelial cells, hydrolyzes VLDL triglycerides and liberates free fatty acids for peripheral tissue needs⁷. With the progressive loss of triglycerides VLDL is transformed into LDL (density, 1.019-1.063 g/ml), which represents the main cholesterol carrying protein in plasma (about 60% of total cholesterol). Each LDL particle has a central lipophilic core composed by cholesteryl esters and triglycerides, and an outer shell of phospholipids and free cholesterol⁸. The apolipoprotein (apo) B, which intercalates the LDL surface and holds about 20% of the total volume⁸, acts as a ligand for the receptor that allows cholesterol influx⁵. Cells use cholesterol for building membranes and for synthesizing steroid hormones in adrenal glands.

The B receptor-mediated cholesterol uptake by cells is fine tuned by feedback signals finalized to the maintenance of constant cholesterol levels for cellular requirements⁵. After internalization the LDL particle is fragmented and cholesterol enters the intracellular cholesterol pool. The increase

in intracellular cholesterol concentration suppresses the synthesis of B100 receptors and of the HMG-CoA reductase enzyme. Consequently, cholesterol influx and endogenous cholesterol synthesis are blocked. On the other hand, cholesterol influx activates the acyl-coenzyme A: cholesterol acyltransferase enzyme, which holds down free cholesterol levels⁵. In fact, the excess of free cholesterol is cytotoxic because it forms insoluble crystals. Cholesterol esterification and storage into a pool of inert cytoplasmic cholesterol esters prevent toxicity.

The receptor for oxidized low-density lipoproteins

Circulating LDL are also termed "native LDL" to distinguish them from post-translational modified LDL, which enhance its uptake by macrophage *in vitro*. Modifications of LDL include self-aggregation, complexation with proteoglycans, immunoglobulins, and degradation by hydrolytic enzymes⁹⁻¹⁷. However, none of these modifications has been extensively studied as LDL modification based on oxidation. The papers dealing with oxidized LDL increased exponentially in the last two decades. Oxidized LDL can be produced by several physiological plausible mechanisms that give to oxidation a central role in the theory of atherogenesis^{1,5}. Numerous studies have supported the occurrence of oxidized LDL *in vivo*. Oxidized LDL, by-products, and epitopes of LDL oxidation have extensively been demonstrated in atherosclerotic plaques¹⁸⁻²⁴, and even small amounts of oxidized LDL can be demonstrated in plasma²⁵⁻²⁹. The discovery of oxidized LDL was coincident with studies of cellular uptake of cholesterol⁵. It was evident that not all cell types use the native LDL receptor for cholesterol influx. Macrophages, the main cellular type of atherosclerotic plaque, do not express classic LDL receptors and do not accumulate native LDL at appreciable rate. Initially, it was shown that derivatization of apo B by acetylation of lysine residues rendered LDL suitable for influx into macrophages⁶. Thus, macrophages express a different receptor that is not down-regulated by cholesterol influx, and engulf lipids until becoming foam cells. However, it was not plausible that acetylation of apo B could have occurred *in vivo*. In searching a physiologic mechanism for apo B derivatization, by-products of oxidation of LDL lipids resulted effective derivatizers of apo B. It was shown that LDL incubated in the presence of transition metals induced modification of LDL suitable for macrophage uptake comparable to that of acetylated LDL. The modification of LDL induced by transition metal ions is the result of an oxidation process. Lipid peroxidation is one of the deleterious effects of free radical attack to organic molecules of living organisms. Among lipids, cholesterol and polyunsaturated fatty acids (PUFA) are very sensitive to free radical attack. PUFA are easily attacked at the

level of α -methylene hydrogens with consequent rearrangement, formation of a lipid radical, and triggering of a chain reaction with incorporation of molecular oxygen⁸. By-products of lipid peroxidation, including aldehydes – the most popular being malondialdehyde – and ketones, have enough chemical reactivity to derivatize apo B and to conjugate to other lipids⁸. In addition apo B can be directly modified by oxidative damage³⁰.

Oxidized LDL exhibit the following chemical properties: increased electrophoretic mobility, increased lisophosphatides, decreased phosphatidylcholine, increased conjugated dienes, hydroperoxides, oxysterols and isoprostanes, and depletion of constitutive antioxidants⁸.

Free radical-mediated LDL oxidation has also been produced by physiological plausible mechanisms based on enzymes – lipoxygenase and phospholipase A₂³¹, redox-cycling of hemoglobin³², and by cells³³⁻³⁵.

The original acetyl LDL receptor has been cloned and now designated scavenger receptor A (SRA) and it occurs in two differentially spliced forms, SRAI and SRAII, which have comparable ligand binding capacity^{36,37}. The pathogenic role of SRA in atherogenesis has been demonstrated by crossing SRA-targeted mice with apo E-targeted and finding reduction of atherosclerosis lesion severity³⁸.

The B class of scavenger receptors includes CD36 and SRB1^{39,40}. Macrosialin and CD68 have a partial oxidized LDL binding capacity^{41,42}. However, taking into account that these receptors are little expressed on plasma membrane, their role in the uptake of oxidized LDL is uncertain.

Pro-atherogenic role of oxidized low-density lipoproteins

Oxidized LDL possess several biological activities, other than the affinity for the scavenger receptor, relevant for atherogenesis. Oxidized LDL induce endothelial dysfunction and the expression of adhesion molecules on endothelial cells^{43,44}, favor monocyte differentiation⁴⁵, are cytotoxic for most cells, counteract biological effects of nitric oxide⁴⁶, and are thrombogenic (through platelet activation and increase in tissue factor activity from endothelial cells)^{47,48}.

Oxidized LDL can stimulate the release of macrophage colony-stimulating factor and monocyte chemoattractant protein-1 by endothelial cells with a potential role in facilitating the development of fatty streaks by recruiting monocytes and facilitating their differentiation into macrophages^{49,50}.

Isoprostanes, which are generated during LDL oxidation and found in the cellular component of atherosclerotic lesions, exhibit mitogenic activity and induce platelet aggregation and vasoconstriction⁵¹. In analogy, cholesterol oxidation products have also shown biolog-

ical activities relevant in the atherosclerosis process. 7β -hydroperoxy-5-en-3 β -ol, generated during LDL oxidation, is a component of atheroma, is cytotoxic^{21,52} and is implicated in the expression of several cytokines, which are of potential relevance in atherosclerosis⁵²⁻⁵⁶.

Antioxidants and atherosclerosis

Lipids in LDL are protected from free radical-mediated oxidation by several lipophilic antioxidants. On molar basis, the most represented antioxidant is α -tocopherol (11 nmol/mg protein, equal to 6 molecules α -tocopherol in an LDL particle). Gamma-tocopherol, ubiquinol-10, carotenoids and other antioxidants are present in much smaller amounts⁸.

Antioxidants prevent LDL oxidation *in vitro* and the biological effects of oxidized LDL, including macrophage lipid loading and cytotoxicity. Thus, antioxidants have been considered as candidates for inhibiting the evolution of atherosclerotic disease¹. This hypothesis has also been supported by studies in several experimental animal models in which the administration of antioxidants slows the progression of atherosclerosis^{57,58}. Vitamin E and synthetic antioxidants (probucol, butylated hydroxytoluene and diphenylene-diamine) have been shown to inhibit the progression of the disease in the LDL receptor-deficient rabbit and mouse, in apo E-deficient mouse, in cholesterol-fed hamster, New Zealand White rabbit and cynomolgus monkey^{1,58}.

Markers of low-density lipoprotein oxidation and coronary heart disease

In the recent years new technologies have been developed to screen for the existence of enhanced oxidant stress in humans. Isoprostanes are a new family of eicosanoids that derive from oxidation of arachidonic acid⁵¹. They have been found elevated in atherosclerotic plaque and in several settings associated with coronary atherosclerosis. In particular, isoprostanes have been shown to be elevated in hypercholesterolemic and diabetic patients and in patients undergoing coronary angioplasty⁵⁹⁻⁶¹. An open issue is if isoprostanes are mere markers of oxidant stress or have also biological relevance in the context of atherosclerosis and its complications. Isoprostanes, for instance, are not able to induce platelet aggregation, but in a range of concentration between 10 nM and 10 μ M enhance platelet response to subthreshold concentration of different agonists, such as adenosine diphosphate and collagen⁶². These concentrations are higher than those found in conditions characterized by enhanced oxidant stress. However, it has been postulated that they could have a role in case of coincidence of enhanced platelet aggre-

gation and oxidant stress, as in diabetic and hypercholesterolemic patients. A significant correlation was in fact demonstrated between isoprostanes and urinary thromboxane B₂ in this setting^{59,60}, but a clear-cut cause-effect relationship is still lacking. Another biological effect exerted by isoprostanes is vasoconstriction but also in this case concentrations necessary to lead to this effect are higher than those present in human circulation⁶³. Further studies are, therefore, necessary to better understand the role of isoprostanes in the pathophysiology of atherosclerosis.

Cholesterol oxidation products (oxysterols) represent another possible approach to study oxidant stress in humans. Oxysterols derive from enzymatic and non-enzymatic oxidation of cholesterol and are quite abundant in human circulation and in atherosclerotic plaques⁵². Oxysterols may be useful not only as markers of oxidant stress, but also to identify new markers of endothelial damage in the setting of atherosclerosis. 7β -hydroperoxy-5-en-3 β -ol, a component of atheroma that is produced during LDL oxidation, is cytotoxic and has been implicated in the expression of several cytokines involved in the atherosclerotic process²¹. Oxysterols may represent an intriguing approach to check for atherosclerosis and its complications also because of a slow elimination from human circulation. In fact, oxysterols are formed in minimally modified LDL, which are not recognized by the scavenger receptor. This could be the reason why the plasma concentration of oxysterols is 3 times higher than that of isoprostanes.

Antibodies against oxidized LDL is another approach that has been largely used to detect atherosclerosis progression, and more recently to screen for chronic and acute coronary heart disease. Antibodies against LDL may be directed against minimally modified LDL or fully oxidized LDL. While minimally modified LDL represent the oxidation of PUFA on phospholipids and cholesterol esters, fully oxidized LDL represent the depletion of antioxidant and apo B-derivatization⁶⁴. It is still uncertain if using one of these two antibodies allows a better interpretation of atherosclerosis progression. Actually, there are conflicting results in the sensitivity of these antibodies in identifying patients with acute coronary syndrome.

Oxidized low-density lipoproteins, antioxidants, and cardiovascular disease

The reduction of atherosclerosis lesions in experimental animal models provides a strong support for using antioxidants to prevent and/or reduce the progression of atherosclerosis in man⁵⁸. The oxidized LDL route in plaque resident foam cells and its modulation by vitamin E have been demonstrated *in vivo* in man, in patients undergoing carotid endarterectomy⁶⁵. Radiolabeled LDL intravenously injected have been localized in foam cells of the carotid specimen obtained from en-

darterectomy. The uptake of LDL was almost completely suppressed in foam cells of patients treated for 4 weeks with vitamin E (900 mg daily)⁶⁵.

In aggregate, the impact of oxidation on vascular disease provides a strong conceptual basis for using antioxidants at clinical level. It should be reminded that additional support comes from observational studies that showed that dietary intake of vitamin E is inversely related to coronary heart disease^{57,58}.

Several clinical trials have been conducted with vitamin E to assess if an antioxidant treatment was able to reduce cardiovascular events in patients with previous cardiovascular events or at risk of developing them. Initially, the CHAOS trial showed a significant reduction of cardiovascular events in patients supplemented with vitamin E⁶⁶. However, the lack of effect of vitamin E in subsequent trials raised some concerns on the efficacy of vitamin E in the clinical setting of coronary heart disease^{67,68}. The cause of these conflicting results is not clear and several factors have been suggested to play a role. These include: compliance, the trial design, the dosage regimen used – ranging from 50 to about 1600 mg (equivalent to 800 IU) a day – and the source of vitamin E linked to the different bioavailability of natural and synthetic forms of vitamin E^{57,58,69}. Compliance is questioned because most studies did not estimate vitamin E plasma levels in placebo and target groups⁵⁸. Plasma vitamin E concentration detected in the ATBC study, which used 50 mg synthetic vitamin E daily, was similar to that observed in the CHAOS study, which used 400 or 800 IU natural vitamin E daily⁵⁸.

The dosage regimen is likely of relevant importance in clinical trials as recently supported by the SPACE trial in which 800 IU vitamin E daily significantly reduced vascular events in hemodialysis patients, who are at high risk of cardiovascular complications⁷⁰. Conversely, 300 mg (equivalent to 150 IU) vitamin E daily has been reported to be insufficient to reduce cardiovascular events in patients at risk⁷¹. Therefore, we should be certain that these trials analyzed subjects in whom the plasma antioxidant capacity was increased by vitamin E treatment. Measurement of plasma vitamin E might be an indirect but useful way to study this issue. In addition, plasma vitamin E should be expressed in relation to cholesterol or blood lipids more than simply indicating its plasma levels⁷².

An additional point emerging from clinical trials is that vitamin E bioavailability is never mentioned. Vitamin E is absorbed in the gastrointestinal tract in conjunction with lipids. We have recently demonstrated that in about 30% of subjects vitamin E is not absorbed, if not taken with meal, and that its availability may not be adequate to increase plasma antioxidant activity⁷³. Thus, no change in total antioxidant activity was detected despite plasma vitamin E increased by 29%, suggesting the existence of a cut-off above which changes in the antioxidant activity are observed⁷³.

Conclusions

LDL oxidation and the scavenger receptor are emerged as principal actors in the process of atherosclerosis. Their connections have been deeply investigated *in vitro* and in animal models and the data accumulated constitute the background for understanding atherosclerotic vascular disease. Antioxidants are conceptually the best candidates to interfere with the process and among them vitamin E, as a natural non-toxic compound, has received much attention. However, the intense research in the topic of LDL oxidation has unanswered the question of the relevance of LDL oxidation at clinical level. Clinical trials did not include biochemical markers of oxidant stress for selecting patients who potentially would benefit from antioxidant treatment, nor compliance of antioxidant treatment was available. Until recently, commonly employed indices of lipid peroxidation were based on *ex vivo* systems or unspecific analytes. Accordingly, much caution should be used in the interpretation of data based on unspecific assays for LDL oxidation. The availability of specific and sensitive markers of lipid peroxidation, including isoprostanes and oxysterols, measurable by mass spectrometry should be widely used at clinical level to identify patients with increased oxidant stress. In addition, antioxidant treatment should be monitored by measuring plasma concentrations of the supplemented vitamin, and eventually the changes in plasma antioxidant activity.

References

- Steinberg D. Low density lipoprotein oxidation and its pathobiological significance. *J Biol Chem* 1997; 272: 20963-6.
- Gerrity RG. The role of the monocyte in atherogenesis: I. Transition of blood-borne monocytes into foam cells in fatty lesions. *Am J Pathol* 1981; 103: 181-90.
- Gerrity RG. The role of the monocyte in atherogenesis: II. Migration of foam cells from atherosclerotic lesions. *Am J Pathol* 1981; 103: 191-200.
- Ross R, Glomset JA. The pathogenesis of atherosclerosis (part I). *N Engl J Med* 1976; 295: 369-77.
- Brown MS, Goldstein JL. Lipoprotein metabolism in the macrophage: implications for cholesterol deposition in atherosclerosis. *Annu Rev Biochem* 1983; 52: 223-61.
- Goldstein JL, Ho YK, Basu SK, Brown MS. Binding site on macrophages that mediates uptake and degradation of acetylated low density lipoprotein, producing massive cholesterol deposition. *Proc Natl Acad Sci USA* 1979; 76: 333-7.
- Havel RJ, Kane JP. Introduction, structure and metabolism of plasma lipoproteins. In: Scriver CR, Baudet AL, Sly WS, Valle D, eds. *The metabolic and molecular bases of inherited disease*. Vol II. New York, NY: McGraw-Hill, 1995: 1841-51.
- Esterbauer H, Gebicki J, Puhl H, Jurgens G. The role of lipid peroxidation and antioxidants in oxidative modification of LDL. *Free Radic Biol Med* 1992; 13: 341-90.
- Khoo JC, Miller E, McLoughlin P, Steinberg D. Enhanced macrophage uptake of low density lipoprotein after self-aggregation. *Arteriosclerosis* 1998; 8: 348-58.

10. Hurt E, Camejo G. Effect of arterial proteoglycans on the interaction of LDL with human monocyte-derived macrophages. *Atherosclerosis* 1987; 67: 115-26.
11. Klimov AN, Denisenko AD, Popov AV, et al. Lipoprotein-antibody immune complexes. Their catabolism and role in foam cell formation. *Atherosclerosis* 1985; 58: 1-15.
12. Griffith RL, Virella GT, Stevenson HC, Lopes-Virella MF. Low density lipoprotein metabolism by human macrophages activated with low density lipoprotein immune complexes. A possible mechanism of foam cell formation. *J Exp Med* 1988; 168: 1041-59.
13. Khoo JC, Miller E, Pio F, Steinberg D, Witztum JL. Monoclonal antibodies against LDL further enhance macrophage uptake of LDL aggregates. *Arterioscler Thromb* 1992; 12: 1258-66.
14. Heinecke JW, Suits AG, Aviram M, Chait A. Phagocytosis of lipase-aggregated low density lipoprotein promotes macrophage foam cell formation. Sequential morphological and biochemical events. *Arterioscler Thromb* 1991; 11: 1643-51.
15. Khoo JC, Miller E, McLoughlin P, Steinberg D. Prevention of low density lipoprotein aggregation by high density lipoprotein or apolipoprotein A-I. *J Lipid Res* 1990; 31: 645-52.
16. Bhakdi S, Dorweiler B, Kirchmann R, et al. On the pathogenesis of atherosclerosis: enzymatic transformation of human low density lipoprotein to an atherogenic moiety. *J Exp Med* 1995; 182: 1959-71.
17. Suits AG, Chait A, Aviram M, Heinecke JW. Phagocytosis of aggregated lipoprotein by macrophages: low density lipoprotein receptor-dependent foam-cell formation. *Proc Natl Acad Sci USA* 1989; 86: 2713-7.
18. Palinski W, Rosenfeld ME, Yla-Herttuala S, et al. Low density lipoprotein undergoes oxidative modification in vivo. *Proc Natl Acad Sci USA* 1989; 86: 1372-6.
19. Yla-Herttuala S, Palinski W, Rosenfeld ME, et al. Evidence for the presence of oxidatively modified low density lipoprotein in atherosclerotic lesions of rabbit and man. *J Clin Invest* 1989; 84: 1086-95.
20. Yla-Herttuala S, Palinski W, Rosenfeld ME, Steinberg D, Witztum JL. Lipoproteins in normal and atherosclerotic aorta. *Eur Heart J* 1990; 11 (Suppl E): 88-99.
21. Chisolm GM, Ma G, Irwin KC, et al. 7 beta-hydroperoxycholest-5-en-3 beta-ol, a component of human atherosclerotic lesions, is the primary cytotoxin of oxidized human low density lipoprotein. *Proc Natl Acad Sci USA* 1994; 91: 11452-6.
22. Praticò D, Iuliano L, Mauriello A, et al. Localization of distinct F2-isoprostanes in human atherosclerotic lesions. *J Clin Invest* 1997; 100: 2028-34.
23. Palinski W, Ord VA, Plump AS, Breslow JL, Steinberg D, Witztum JL. ApoE-deficient mice are a model of lipoprotein oxidation in atherogenesis. Demonstration of oxidation-specific epitopes in lesions and high titers of autoantibodies to malondialdehyde-lysine in serum. *Arterioscler Thromb* 1994; 14: 605-16.
24. Palinski W, Tangirala RK, Miller E, Young SG, Witztum JL. Increased autoantibody titers against epitopes of oxidized LDL in LDL receptor-deficient mice with increased atherosclerosis. *Arterioscler Thromb Vasc Biol* 1995; 15: 1569-76.
25. Palinski W, Horkko S, Miller E, et al. Cloning of monoclonal autoantibodies to epitopes of oxidized lipoproteins from apolipoprotein E-deficient mice. Demonstration of epitopes of oxidized low density lipoprotein in human plasma. *J Clin Invest* 1996; 98: 800-14.
26. Itabe H, Yamamoto H, Imanaka T, et al. Sensitive detection of oxidatively modified low density lipoprotein using a monoclonal antibody. *J Lipid Res* 1996; 37: 45-53.
27. Tamai O, Matsuoka H, Itabe H, Wada Y, Kohno K, Imaizumi T. Single LDL apheresis improves endothelium-dependent vasodilation in hypercholesterolemic humans. *Circulation* 1997; 95: 76-82.
28. Holvoet P, Theilmeier G, Shivalkar B, Flameng W, Collen D. LDL hypercholesterolemia is associated with accumulation of oxidized LDL, atherosclerotic plaque growth, and compensatory vessel enlargement in coronary arteries of miniature pigs. *Arterioscler Thromb Vasc Biol* 1998; 18: 415-22.
29. Sevanian A, Bittolo-Bon G, Cazzolato G, et al. LDL- is a lipid hydroperoxide-enriched circulating lipoprotein. *J Lipid Res* 1997; 38: 419-28.
30. Berlett BS, Stadtman ER. Protein oxidation in aging, disease, and oxidative stress. *J Biol Chem* 1997; 272: 20313-6.
31. Neuzil J, Upston JM, Witting PK, Scott KF, Stocker R. Secretory phospholipase A₂ and lipoprotein lipase enhance 15-lipoxygenase-induced enzymic and nonenzymic lipid peroxidation in low-density lipoproteins. *Biochemistry* 1998; 37: 9203-10.
32. Miller YI, Altamentova SM, Shaklai N. Oxidation of low-density lipoprotein by hemoglobin stems from a heme-initiated globin radical: antioxidant role of haptoglobin. *Biochemistry* 1997; 36: 12189-98.
33. Aviram M. LDL-platelet interaction under oxidative stress induces macrophage foam cell formation. *Thromb Haemost* 1995; 74: 560-4.
34. Henriksen T, Mahoney EM, Steinberg D. Enhanced macrophage degradation of low density lipoprotein previously incubated with cultured endothelial cells: recognition by receptors for acetylated low density lipoproteins. *Proc Natl Acad Sci USA* 1981; 78: 6499-503.
35. Cathcart MK, Morel DW, Chisolm GM III. Monocytes and neutrophils oxidize low density lipoprotein making it cytotoxic. *J Leukoc Biol* 1985; 38: 341-50.
36. Krieger M, Herz J. Structures and functions of multiligand lipoprotein receptors: macrophage scavenger receptors and LDL receptor-related protein (LRP). *Annu Rev Biochem* 1994; 63: 601-37.
37. Krieger M, Acton S, Ashkenas J, Pearson A, Penman M, Resnick D. Molecular flypaper, host defense, and atherosclerosis. Structure, binding properties, and functions of macrophage scavenger receptors. *J Biol Chem* 1993; 268: 4569-72.
38. Suzuki H, Kurihara Y, Takeya M, et al. A role for macrophage scavenger receptors in atherosclerosis and susceptibility to infection. *Nature* 1997; 386: 292-6.
39. Endemann G, Stanton LW, Madden KS, Bryant CM, White RT, Proctor AA. CD36 is a receptor for oxidized low density lipoprotein. *J Biol Chem* 1993; 268: 11811-6.
40. Acton SL, Scherer PE, Lodish HF, Krieger M. Expression cloning of SR-BI, a CD36-related class B scavenger receptor. *J Biol Chem* 1994; 269: 21003-9.
41. Ramprasad MP, Fischer W, Witztum JL, Sambrano GR, Quehenberger O, Steinberg D. The 94- to 97-kDa mouse macrophage membrane protein that recognizes oxidized low density lipoprotein and phosphatidylserine-rich liposomes is identical to macrosialin, the mouse homologue of human CD68. *Proc Natl Acad Sci USA* 1995; 92: 9580-4.
42. Ramprasad MP, Terpstra V, Kondratenko N, Quehenberger O, Steinberg D. Cell surface expression of mouse macrosialin and human CD68 and their role as macrophage receptors for oxidized low density lipoprotein. *Proc Natl Acad Sci USA* 1996; 93: 14833-8.
43. Kugiyama K, Kerns SA, Morrisett JD, Roberts R, Henry PD. Impairment of endothelium-dependent arterial relaxation by lysolecithin in modified low-density lipoproteins. *Nature* 1990; 344: 160-2.

44. Frostegard J, Haegerstrand A, Gidlund M, Nilsson J. Biologically modified LDL increases the adhesive properties of endothelial cells. *Atherosclerosis* 1991; 90: 119-26.
45. Frostegard J, Nilsson J, Haegerstrand A, Hamsten A, Wigzell H, Gidlund M. Oxidized low density lipoprotein induces differentiation and adhesion of human monocytes and the monocytic cell line U937. *Proc Natl Acad Sci USA* 1990; 87: 904-8.
46. Schmidt K, Graier WF, Kostner GM, Mayer B, Kukovetz WR. Activation of soluble guanylate cyclase by nitrovasodilators is inhibited by oxidized low-density lipoprotein. *Biochem Biophys Res Commun* 1990; 172: 614-9.
47. Weis JR, Pitas RE, Wilson BD, Rodgers GM. Oxidized low-density lipoprotein increases cultured human endothelial cell tissue factor activity and reduces protein C activation. *FASEB J* 1991; 5: 2459-65.
48. Volf I, Roth A, Cooper J, Moeslinger T, Koller E. Hypochlorite modified LDL are a stronger agonist for platelets than copper oxidized LDL. *FEBS Lett* 2000; 483: 155-9.
49. Cushing SD, Berliner JA, Valente AJ, et al. Minimally modified low density lipoprotein induces monocyte chemotactic protein 1 in human endothelial cells and smooth muscle cells. *Proc Natl Acad Sci USA* 1990; 87: 5134-8.
50. Rajavashist TB, Andalibi A, Territo MC, et al. Induction of endothelial cell expression of granulocyte and macrophage colony-stimulating factors by modified low-density lipoproteins. *Nature* 1990; 344: 254-7.
51. Praticò D. F(2)-isoprostanes: sensitive and specific non-invasive indices of lipid peroxidation in vivo. *Atherosclerosis* 1999; 147: 1-10.
52. Brown AJ, Jessup W. Oxysterols and atherosclerosis. *Atherosclerosis* 1999; 42: 1-28.
53. Liu Y, Hulten LM, Wiklund O. Macrophages isolated from human atherosclerotic plaques produce IL-8, and oxysterols may have a regulatory function for IL-8 production. *Arterioscler Thromb Vasc Biol* 1997; 17: 317-23.
54. Jovinge S, Ares MP, Kallin B, Nilsson J. Human monocytes/macrophages release TNF-alpha in response to Ox-LDL. *Arterioscler Thromb Vasc Biol* 1996; 16: 1573-9.
55. Ku G, Thomas CE, Akeson AL, Jackson RL. Induction of interleukin-1 beta expression from human peripheral blood monocyte-derived macrophages by 9-hydroxyoctadecadienoic acid. *J Biol Chem* 1992; 267: 14183-8.
56. Malden LT, Chait A, Raines EW, Ross R. The influence of oxidatively modified low density lipoproteins on expression of platelet-derived growth factor by human monocyte-derived macrophages. *J Biol Chem* 1991; 266: 13901-7.
57. Pryor WA. Vitamin E and heart disease: basic science to clinical intervention trials. *Free Radic Biol Med* 2000; 28: 141-64.
58. Violi F, Micheletta F, Iuliano L. Vitamin E, atherosclerosis and thrombosis. *Thromb Haemostas* 2001; 85: 766-70.
59. Davì G, Alessandrini P, Mezzetti A, et al. In vivo formation of 8-epi-prostaglandin F_{2α} is increased in hypercholesterolemia. *Arterioscler Thromb Vasc Biol* 1997; 17: 3230-5.
60. Davì G, Ciabattoni G, Consoli A, et al. In vivo formation of 8-iso-prostaglandin F_{2α} and platelet activation in diabetes mellitus. Effects of improved metabolic control and vitamin E supplementation. *Circulation* 1999; 99: 224-9.
61. Iuliano L, Praticò D, Greco C, et al. Angioplasty increases coronary sinus F2-isoprostane formation: evidence for in vivo oxidative stress during PTCA. *J Am Coll Cardiol* 2001; 37: 76-80.
62. Praticò D, Smyth EM, Violi F, FitzGerald GA. Local amplification of platelet function by 8-epi prostaglandin F2alpha is not mediated by thromboxane receptor isoforms. *J Biol Chem* 1996; 271: 14916-24.
63. Cracowski JL, Devillier P, Durand T, Stanke-Labesque F, Bessard G. Vascular biology of the isoprostanes. *J Vasc Res* 2001; 38: 93-103.
64. Tsimikas S, Witztum JL. Measuring circulating oxidized low-density lipoprotein to evaluate coronary risk. *Circulation* 2001; 103: 1930-2.
65. Iuliano L, Mauriello A, Sbarigia E, Spagnoli LG, Violi F. Radiolabeled native low-density lipoprotein injected into patients with carotid stenosis accumulates in macrophages of atherosclerotic plaque: effect of vitamin E supplementation. *Circulation* 2000; 101: 1249-54.
66. Stephens NG, Parsons A, Schofield PM, et al. Randomised controlled trial of vitamin E in patients with coronary disease: Cambridge Heart Antioxidant Study. *Lancet* 1996; 347: 781-6.
67. The Heart Outcomes Prevention Evaluation Study Investigators. Vitamin E supplementation and cardiovascular events in high-risk patients. *N Engl J Med* 2000; 342: 154-60.
68. GISSI-Prevenzione Investigators. Dietary supplementation with n-3 polyunsaturated fatty acids and vitamin E after myocardial infarction: results of the GISSI-Prevenzione trial. *Lancet* 1999; 354: 447-55.
69. Jialal I, Devaraj S. Vitamin E supplementation and cardiovascular events in high-risk patients. *N Engl J Med* 2000; 342: 1917-8.
70. Boaz M, Smetana S, Weinstein T, et al. Secondary prevention with antioxidants of cardiovascular disease in endstage renal disease (SPACE): randomised placebo-controlled trial. *Lancet* 2000; 356: 1213-8.
71. Low-dose aspirin and vitamin E in people at cardiovascular risk: a randomised trial in general practice. Collaborative Group of the Primary Prevention Project. *Lancet* 2001; 357: 89-95.
72. Traber MG, Jialal I. Measurement of lipid-soluble vitamins: further adjustment needed? *Lancet* 2000; 355: 2013-4.
73. Iuliano L, Micheletta F, Maranghi M, Frati G, Diczfalusy U, Violi F. Bioavailability of vitamin E as function of food intake in healthy subjects. Effects on plasma peroxide-scavenging activity and cholesterol-oxidation products. *Arterioscler Thromb Vasc Biol*, in press.